TABLE 1.-Continued.

Date, Localities, Soil.	Geological Forma- tion of Soil.	Azoto- bacter p.g. of Soil.	Beijer- inckia p.g. of Soil.	рН.	P. (1) (2) Mg./kg. Mg./kg.		C. G./kg. of Soil.
(46) 29.7.52. 20 miles sonth of Rock- hampton. Alluvial soil adjoining dam. Open semi-cultivated land		_		5.8			25.0
with light timber. (47) 29.7.52. 20 miles north of Glad- stone. Brown shale. Coastal open forest with grass.	wine	-		5.7	_	—	20.0
(48) 27.7.52. 40 miles north of Clare- mont. Black (?) alluvial soil. Dry sclerophyll forest.	-	-	_	5.75	_	—	5.5

The soil samples were collected from that portion of the route from the eastern Kimberleys, north-west coast, selerophyll woodlands southwards along the Stuart Highway and semi-desert terrain eastwards on the Barkley Highway, western Queensland, to the rain forests of north-cast Queensland.

The geographic distribution of Beijerinckia in Australia seems to be limited at . 17-18° latitude. Of the 15 samples collected around the 19.5° latitude only one gave positive growth of Beijerinckia. It is possible that no Beijerinckia was detected south of 20° latitude. It should be kept in mind that the soil samples examined were in limited number for such a big area. However, if McKnight's results are compiled with the present investigation, it seems that Beijerinckia does not occur south of the Tropic of Capricorn. It is important and very desirable to test more soil samples between 17° and 20° latitude in Australia before a definite conclusion can be made. The limit of 17-18° latitude may be used as a starting point for further investigations.

Authors.			Localities.		% Soil Containing		
					Azotobacter.	Beijerinckia.	
Tensen			New South Wales		25		
McKnight			Queensland		$43 \cdot 15$		
Swaby			Victoria		26.15		
Fchan			Sydney		22	_	
Tchan	·		Northern Australia		15	35	

The ecological conditions of *Beijerinckia* in Australia are not understood. The soil type and its parent material in positive cases are indeed very variable (see Table 1). The analysis of C, P content and pH of soil samples did not show any correlation between presence and absence or number of *Beijerinckia* per g. of soil. On the other hand, *Azotobacter* did not occur at pH less than 5.5 except in one case of sporadic presence of *Azotobacter* at pH = 5.1. The climatic environments have no apparent influence on the presence of *Beijerinckia*.

It is clear that *Beijerinckia* can survive in low rainfall countries. The drought of 1952 in Northern Australia provided an example. Also in high rainfall country outside the tropical zone (Brisbane, Sydney, etc.) *Beijerinckia* seems to be absent (see Text-figure 1). Temperature cannot be considered as an important ecological factor in its distribution, since the occurrence of *Beijerinckia* took place in certain places where the

mean minimum temperature of 57.2° F. (14°C.) is lower than certain places outside the tropic zone, e.g. Brisbane 59.7°F. (15.4°C.).

Under the experimental conditions Beijerinckia (isolated from Northern Australia) inocalated into soil and kept in a refrigerator (+4°C.) survived after 36 days. As along certain parts of the east coast of Australia (Queensland, N.S.W.) the winter temperature never goes much below that limit, there is no apparent reason to believe that Beijerinckiacould be killed during the winter.

Complete desiccation (with CaCl₂) in the laboratory did not destroy *Beijerinckia* in soil after 36 days, but such a severe desiccation is not likely to occur in the temperate places of the east coast of Australia. This may suggest that *Beijerinckia* may survive in air-dried soil at normal humidities.

	Mean Temp	erature (°F.).		Rainfall, (Inches per Year.)
Soil Sample No.	+° max.	+° min.	Humidity.	
32	. 89.4	65 · 3	36	14.72
7	. 92	63 · 9	41	20.84
10, 14, 17, 18, 23, 25, 29 .	. 94	66.7	51	26.48
38, 39, 40	89.9	64.8	52	31.97
41	78	57.2	74	51.84
9, 21	90	74.3	68	60.45
4	82.2	65.5	81	142.61
Bowen	82.6	67.2	69	39.88
Rockhampton	83.5	62.8	67	39.75
Brisbane	78.1	59.7	68	45.27
Sydney	70.2	56.2	70	47.50

TA	BL	E :	з.

(Data compiled from Meteorological data, C.S.I.R.O., Melbourne, 1933. Pamphlet 42.)

If the chemical and meteorological factors could not explain the absence of Beijerinckia outside the tropical zone of Australia, the only remaining hypothesis will be that Beijerinckia is a young genus which may have been introduced to this country very recently and has not had enough time to reach the rest of the continent. This hypothesis may not be considered as a valid one, since the movement of animal and human transport in Queensland is so intensive that the contamination from Northern Queensland can be realized in a matter of months. Furthermore, information in the literature shows that Beijerinckia has been found only in tropical countries. Altson (1936) in Malaya was probably the first person to detect Beijerinckia; later it was found in Indian soils by Starkey and De (1939), in Indonesia and Pacific islands by Derx et al. (1950), in Africa (Kauffmann, 1953) and in South America (Derx, 1952). Outside the tropical zone the Beijerinckia has been sought but unsuccessfully (Northern Africa (Derx, 1952); Southern France (Derx, 1950); N.S.W. (Jensen and Swaby, 1940; Tchan, 1952)). From these data one may conclude that if Beijerinckia has spread so widely in the tropics, including very long distances separated by oceans, it is not likely that it needs more time to reach the Australian soils south of the tropics. It is likely that the hypothesis mentioned above has no important value.

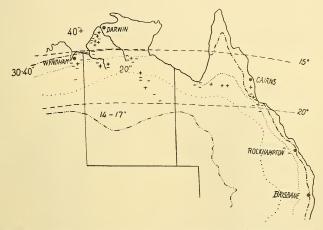
It is not clear why Beijerinckia is a genus confined to tropical countries.

This geographic limitation of *Beijerinckia* can be extended to the distribution of other N-fixing micro-organisms in the world. The distribution of *Rhizobium* is excluded in this paper because it is practically a question of distribution of legumes. The non-symbiotic N-fixing soil micro-organisms can be classified into four groups: blue-green algae (Nostoc and Anabaena), Clostridium, Azotobacter and Beijerinckia.

(1) In the tropical countries the four groups of organisms have been detected.

(2) In the temperate zone the absence of *Beijerinckia* reduces the number to three groups.

(3) In the arctic and antarctic zones the early workers have reported the presence of *Azotobacter*. Rountree (1939) reported the presence of *Azotobacter* in Matquarie Island soils, but more recently Bunt could not confirm this result with soil samples collected in most suitable conditions. Also he reported that on the N-free media the Macquarie Island soil samples gave some colonies similar in appearance to *Azotobacter*.



Text-figure 1.—Map showing distribution of *Beijerinckia*, the latitudes and the rainfall. +..... positive soil; -..... negative soil.

The early positive results may be due to contamination of soil samples. On the other hand, in Greenland, the search for *Azotobacter* has been always negative (Barthel, 1922; Jensen, 1951).

The absence of *Azotobacter* in the very cold regions could be partly explained by the death of *Azotobacter* (including cysts) at a prolonged low temperature. Wang (1949) has reported that *Azotobacter* is killed if the culture is kept in a refrigerator for a prolonged period. So the very cold regions contain only two groups of non-symbiotic N-faxing micro-organisms (blue-green algae and *Clostridium*).

This division of the world, according to the distribution of non-symbiotic N-fixing micro-organisms, into three zones is still at a purely hypothetical stage. It could only be established with some certainty if a very extensive survey in different regions could be carried out. At the present stage this suggestion may provide a starting point for future research work.

CONCLUSION.

The present results have contributed to our knowledge by showing that:

(1) *Beijerinckia* is present in Northern Australia. To the best of my knowledge it is the first time that these organisms have been detected in this country.

(2) The distribution of *Beijerinckia* in Australia seems to be limited to the north of $17-18^{\circ}$ latitude. It is likely to be absent south of the 20° latitude.

(3) The ecological factors of the distribution of *Beijerinckia* in Australia are still not understood. Some chemical, geological and climatic factors are discussed.

(4) A suggestion has been made to divide the world into three zones, according to the distribution of non-symbiotic N-fixing micro-organisms: (a) tropical zone with the presence of *Beijerinckia*, Azotobacter, Clostridium and blue-green algae; (b) temperate zone with Azotobacter, Clostridium and blue-green algae; (c) arctic and antarctic zone with Clostridium and blue-green algae.

This suggestion is purely hypothetical but may provide a starting point for future research.

Acknowledgements.

The author is indebted to Dr. H. S. McKee for his help and criticism; to Mr. H. Fletcher, leader of the Australian Museum Central and North-West Expedition, and particularly to Mr. J. A. Keast, who collected the soil samples; also to Mr. Lovett, of C.S.I.R.O., for sending 15 soil samples from Queensland. This work would have been impossible without their collaboration. His sincere thanks are extended to Dr. W. R. Browne for geological information, to Professor H. G. Derx for his private communications and to Dr. A. B. Walkom for his help.

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EXPLANATION OF PLATE X, FIGS. 3, 4.

Fig. 3.-Photomicrograph of Beijerinckia in mixed culture.

Fig. 4.—Top: Colonies of *Beijerinckia* on N-free medium three weeks old. Lower right: Colonies of *Azotobacter*, same age. Lower left: Colonies of other bacteria.

A NEW SPECIES OF *PSEUDOPHRYNE* FROM VICTORIA. By JOHN A. MOORE, Fulbright Research Scholar, Sydney University.

(One Text-figure.)

[Read 29th July, 1953.]

The Australian Museum has in its collections an undescribed *Pseudophryne* of a most striking kind. It is represented by a single specimen. The basing of a new species on a single specimen is a hazardous procedure, but in this instance I think it is justified in view of the unusual characteristics shown by the specimen. There is no other *Pseudophryne* that has even a remote resemblance.

PSEUDOPHRYNE CORROBOREE, n. sp.

Type: R 13103, a male in the Australian Museum, Sydney. Collected by Ossie Rixon at Towong Hill Station, Corryong, Victoria. Donated by T. W. Mitchell. The type locality is near the Victoria-New South Wales border, about 25 miles north-west of Mount Kosciusko.



Pseudophryne corroboree in dorsal (left) and ventral (right) views. Approximately twice natural size.

Description: A Pseudophryne having the same general structural features as P. australis (Gray) and P. bibroni Günther. Body length 24 mm.; tibia 7.9 mm.; width of head at posterior end of jaws 7.0 mm.; tip of snout to centre of nares 1.1 mm.; centre of nares to anterior corner of eye 1.7 mm.; anterior-posterior dimension of eye 2.2 mm. The three dimensions last given were obtained by viewing the specimen laterally under a binocular microscope and the measurements made with an ocular scale. The fourth toe reaches the snout when the leg is extended along the side of the body. The shape of the head and the structure of the hand and foot are the same as in P. australis and

P. bibroni. A detailed description of these and other members of the genus will be found in Parker (1940).

This species differs from all others of the genus in its unusual dorsal pattern, which can be best appreciated by reference to the figure. In the type the dark bands are black and the light areas pale yellow. Dark and light areas of similar tones cover the entire body. The tubercles at the base of the fingers and on the metacarpals are light, contrasting strongly with the dark background. Many of the tubercles of the foot, including the inner metatarsal tubercle, are likewise light in colour against a dark background. The postfemoral glands cannot be distinguished externally, but the area where they occur in other species of *Pseudophryne* is light in colour.

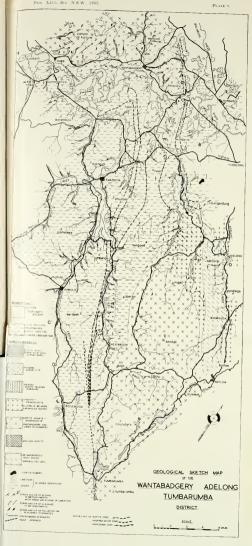
Diagnosis: Pseudophryne corroborce can be distinguished from all other species of the genus and from all other Australian frogs by the boldly contrasting dark and light stripes on the dorsal surface.

The specific name was suggested by the resemblance of the dorsal pattern of *P. corroboree* to the body paintings used by some Australian aboriginal tribes in their corroborees.

I am indebted to Mr. Kinghorn for allowing me to describe this species.

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